BIOLOGY 446 (credit 3 hours) NEUROPHYSIOLOGY LABORATORY Fall 2020-Covid semester

Instructor: ROBIN COOPER, Ph.D.

Office: Biology, room 226
Office phone: 7-5950
E-mail: RLCOOP1@uky.edu

Required Texts

None. Handouts and www based content.

See teaching page: http://web.as.uky.edu/Biology/faculty/cooper/teachingpage.htm

Supplementary Materials

Readings from the primary literature will be assigned on occasion. These articles will be posted on Canvas or the class web page for you to download and read.

Time: Tue - 2:00 to 2:50 PM Lecture; Thu - 1:00 to 3:50 PM Lab Multidisciplinary Science Bldg.

725 Rose Street, Room 155B.

This is the red brick and stucco building directly across the street from Biology (Thomas Hunt Morgan Building), right next to the UK clinic parking garage

Course Description

The course will focus on the experimentation in primarily neurophysiology with some emphasis in muscle physiology. Generation of receptor potentials in sensory neurons will be measured as well as action potentials in axons. Pharmacological experimentation of ionotropic and metabotropic receptors subtypes and second messengers signaling will be conducted. The key role of ion channels and transporters in regulation of the membrane potential will be covered in great detail. The concept of electrochemical equilibrium will be introduced, and quantitative description of the equilibrium membrane potential will include discussion of Goldman and Nernst equations and their applications. The mechanisms of action potential generation, as a result of synaptic and receptor stimulation within a neural cell, will be measured in terms of voltage gated ion channels.

The mechanisms of neuron-neuron communication through electrical and chemical synapses will be examined in live preparations. The function of gap junction ion channels will be examined and measured at electrical synapses. The complex intracellular mechanisms controlling neurotransmitter release at the chemical synapses will be examined. The proteins participating in the SNARE complex, involved in the synaptic vesicle fusion, as well as the molecular machinery involved in synaptic vesicles recycling will be emphasized along with quantal analysis of synaptic transmission. The historical introduction of the quantal hypothesis and its experimental conformation will be covered. Synaptic plasticity will be covered up to the latest discoveries in the field.

Course Objectives

By the end of this course, you should:

1. Have a conceptual understanding of the information processing in the nervous system.

- 2. Understand the molecular mechanisms that enable signal transmission in the nervous system in terms of receptor potentials, synaptic potentials and action potentials.
- 3. Know what the cellular specializations and the molecular machinery are involved in the neuron-neuron communication at the state-of-the-art level.
- 4. Develop a basic knowledge of the sensory processing.
- 5. Be able to understand and critically analyze research papers in the field of Neuroscience.
- 6. Be able to develop new ideas and suggest future research directions in the field of Neuroscience.

The overall objective of the course is to develop insightful understanding of the neurological processes at molecular and cellular level by experimentation. The course will complement other courses offered within the new Neuroscience Major we are establishing on campus.

Pre-requisites are BIO 302 and/or BIO 350 and /or consent of instructor

Course Work/Grading

Overall

Grades will be based on exams, homework assignments, and class participation. There will be two exams. Homework due dates and exam dates are listed in the schedule at the end of this document.

- Class participation: 10% (includes attendance (see notes related to COVID below), adding to the discussion during lectures and journal clubs, and actively participating in in-class exercises such as problem solving and other activities)
- Conducting all laboratory exercises 30% (review of lab notebook, attendance)
- Homework/problem sets: 20%
- Lab report 20% (submitted in a journal publication format and one class presentation -we might have to do this on Zoom as a class)

- Exam 1: 10% - Exam 2: 10%

All exams are required of all students.

There will be classroom presentations in front of the whole class by the students as a means of going over the exams and quizzes as well as presenting your research report. If you have anxiety issues for classroom presentations please see the disability office at UK and obtain an official letter that you need out of giving a classroom presentation, otherwise you are expected to be able to talk in front of the class.

If you are unable to take an examination as scheduled, it is your responsibility to contact me before the exam. There will be make-up exams for excused absences over similar material. Unexcused absences from an exam will result in a score of zero for that test. Homework and problem sets that are turned in late will be

marked down a point each day they are late, and they must be turned in before those that were turned in on time are returned, unless an excused absence. If an excused absence a student will have 1 week to hand in the material.

Quizzes

Quizzes will be assigned to each topic. The quizzes score will be used as an extra credit toward your exam score. Four quizzes within the semester might be given and each is worth 10 points.

Attendance:

Required for all laboratory sessions. See University rules for excused absences University Senate Rule ("SR") 5.2.4.2 (advance notice of an absence must be provided in two circumstances; SR 5.2.4.2.D; SR 5.2.4.2.C.). 5.2.4.2.A excused Absence for Illness.

Format

In this course, you will learn the fundamentals of neural processing through a variety of activities, including lectures, problem sets, independent/group study, and in-class exercises. Please note that you are responsible for all the material in the assigned chapters, including figures, summaries, and "boxes," regardless of whether it is covered in lectures. Thus, you will be responsible for covering some material from the text or readings on your own.

Canvas/Class Communications

Course announcements, assignments, lecture outlines and additional materials will be posted online using Canvas. Exams and homework dates will remain fixed. Updates to this syllabus (regarding topics and reading) will be posted; please check periodically. You will also receive important course announcements via your UK e-mail account. If you do not use your UK e-mail account, you need to activate it. It is strongly recommended that you check your e-mail regularly. I may send messages-sometimes with attachments-to the class using this medium. You should also feel free to e-mail me if you have any questions or problems. Feel free to call me as well, if you prefer a more personal communication. I am also available during the office hours. If you would like to meet with me at another time, please don't hesitate to e-mail or to call, and I can schedule a time to meet.

Honesty and Civility

You must abide by UK's Code of Conduct which prohibits:

- 1. Academic dishonesty and impropriety, including plagiarism and academic cheating.
- 2. Interfering or attempting to interfere with or disrupting the conduct of classes or any other normal or regular activities of the University.

We take plagiarism and other forms of cheating very seriously. If you have any questions as to whether something is plagiarism, please ask me, or, if that's not possible, assume that it is and don't do it!

Disabilities

Any student who needs accommodation because of a disability should contact me privately to discuss the specific situation as soon as possible. The Office of Disability

Resources and Services. They can coordinate reasonable accommodations for students with documented disabilities.

Fall 2020 Academic Calendar

Several modifications to the academic calendar have been made in response to the COVID-19 situation. The current calendar is available https://www.uky.edu/registrar/fall-2020-semester

The calendar features a Reading Day. The current Dead Week restrictions on certain instructional activities would continue to apply to Reading Days. An additional restriction would apply to Reading Days, namely no required class meetings or, more generally, no "required interactions." Reading Days are not academic holidays.

Excused Absences

Academic policies regarding excused absences can be found in the Senate Rules under "Excused Absences." The Senate Council has interpreted excused absences for the Fall 2020 semester to include an excuse from required in-person interactions if the student has been directed to self-quarantine by the University (including its app), a medical professional, public health professional, or government official.

Recommended Syllabus Statements

Face Covering/Distancing Policy

The Senate Council endorses the following recommended syllabus statements for faculty teaching face-to-face courses.

 In accordance with University guidelines, students must wear UK-approved face coverings in the classroom and academic buildings (e.g., faculty offices, laboratories, libraries, performance/design studios, and common study areas where students might congregate). If UK-approved face coverings are not worn over the nose and mouth, students will be asked to leave the classroom.

- Students should complete their daily online wellness screening before accessing university facilities and arriving to class.
- Students should not move chairs or barriers in classrooms and should socially distance at all times, leaving a six (6) foot radius from other people. Masks and hand sanitizer can be found {specific location in building} if needed.
- Students should leave enough space when entering and exiting a room. Students should not crowd doorways at the beginning or end of class.
- The instructor may choose to remove a mask when pedagogically necessary at the front of the classroom and behind a clear barrier. The instructor's mask will be replaced when it is no longer necessary to have it removed, or when the class meeting is complete.

Class Recording Notification

The University of Kentucky Student Code of Conduct defines Invasion of Privacy as using electronic or other devices to make a photographic, audio, or video record of any person without their prior knowledge or consent when such a recording is likely to cause injury or distress.

Meetings of this course may be recorded. All video and audio recordings of lecturers and class meetings, provided by the instructors, are for educational use by students in this class only. They are available only through the Canvas shell for this course and are not to be copied, shared, or redistributed.

As addressed in the Student Code of Conduct, students are expected to follow appropriate university policies and maintain the security of linkblue accounts used to access recorded class materials. Recordings may not be reproduced, shared with those not enrolled in the class, or uploaded to other online environments.

If the instructor or a University of Kentucky office plans any other uses for the recordings, beyond this class, students identifiable in the recordings will be notified to request consent

prior to such use. In anticipation of such cases, students may be asked to complete an "authorization of use" form by a faculty member.

Video and audio recordings by students are not permitted during the class unless the student has received prior permission from the instructor. Any sharing, distribution, and or uploading of these recordings outside of the parameters of the class is prohibited. Students with specific recording accommodations approved by the Disability Resource Center should present their official documentation to the instructor.

All content for this course, including handouts, assignments, and lectures are the intellectual property of the instructors and cannot be reproduced or sold without prior permission from the instructors. A student may use the material for reasonable educational and professional purposes extending beyond this class, such as studying for a comprehensive or qualifying examination in a degree program, preparing for a professional or certification examination, or to assist in fulfilling responsibilities at a job or internship.

TOPICS AND READINGS—TENTATIVE SCHEDULE

Please note that this is a tentative schedule and may be modified depending on how the course is progressing. All changes will be announced in advance, and students will be well aware of them—particularly regarding what is going to be included on exams.

Week, Topics, Student-driven In-class Activities. *The schedule will change depend on the availability of animals and the COVID-19 changes.*

- Week 1. Learn about equipment (extracellular & intracellular amps, microscopes, electrode puller). Solutions and laboratory tools. Animal care. Lab notebooks & reports.
- Week 1 & 2. We will start on the bread board to learn about OHM's law and electrical conduction(see this lab Page- Neuron model)
- Week 2 & 3. Earthworm preparation to learn about conduction velocity and refractory periods(see this lab Page- <u>Earth worm experiments</u>)

Weeks (maybe week 2) 3 & 4. - Measure membrane potentials in crayfish abdomen muscles and plot Rp vs [Na]o graphs and Rp vs [K]o graphs. (see this lab PAGE- RP, K, Na)

- Weeks 3 &4 . Measure facilitation and depression in tonic and phasic neuromuscular junctions in crayfish abdomen muscles. We will learn how to stimulate motor nerves and record EPSPs/IPSPs. (see this lab PAGE- EPSPs)
- Week 5. Learn to record from proprioceptors (extracellular) in the crab leg and relate to joint positions. Research paper discussion. (see this lab PAGE- <u>Joint receptors in a crab</u>)
- Week 6. Learn how to forward fill neurons from the crab leg proprioceptors (CoCl2, 4-Di-2 ASP) as well as stain with methylene blue. (see this lab PAGE- <u>Staining of Joint</u> receptors in a crab)
- Week 7. Review EXAM 1 Learn to record from tension receptors in the crab leg related to muscle length and contraction. (see this lab PAGE- tension nerve recordings)
- Week 7. EEG Human lab (go to lab EEG lab page)
- Week 8. Introduction to the leech nervous system and practice dissection. Staining of leech ganglia.
- Week 8. Learn how to dissect the leech ventral nerve cord and obtain intracellular recordings from identified neurons. Current injections and threshold measures. Potentially two intracellular electrodes and record in situ synaptic connections. Investigate the ionic currents making up the action potentials. (see this lab PAGE-Leech ganglion lab)
- Week 8 & 9. -Mapping skin receptive fields on the leech while recording from neurons. Dye fills. (see this lab PAGE- Leech Skin lab)
- Spring break
- Week 10. Learn how to remove and culture leech neurons for forming synapses in culture. (see this lab PAGE- <u>Leech neuron culture lab</u>)
- Week 11. -Catch up on experiments and analysis day. Graph data, catch up on homework and literature gathering. Plot I V curves and use Ohms law to determine Rm with crayfish skeletal muscle.
- Week 11. Quantal analysis of synaptic transmission: Crayfish NMJ record quantal responses..(see this lab PAGE- <u>quantal analysis</u>)
- Week 12. Sensory recordings from the cockroach cerci. (see this lab PAGE-<u>cockroach</u> cerci)
- Week 13. Sensory recordings from the cockroach cerci. (see this lab PAGE-<u>cockroach</u> cerci)

Week 13. Vision: crayfish caudal photo receptor in crayfish.

.....(see this lab PAGE- Crayfish photoreceptors lab)

Week 14. Work on your own out of lab for write ups. Finish off class projects.

Week 15. - Make up any labs needed. Make sure lab report is going along OK. Pass out take home exam.(Exam 2)

Week 16. (Finals week) Turn in final exam and reports.